

# DATA PROCESSING IN GENOMICS: FROM THE SEQUENCERS TO THE RESULTS

GIGA Doctoral School for Health Sciences  
Introduction to scientific computing

ARNAUD LAVERGNE

GIGA-Genomics

Bioinformatic team



# GIGA - GENOMICS

- Platform
  - Sequencing services
    - Biological materials
    - DNA/RNA
    - Libraries
    - Sequencers
    - Data
  - High Throughput Sequencing (HTS) / Next Generation Sequencing (NGS)
  - Bioinformatic team
    - Data analysis
    - And more ...



Since the development of new next generation sequencing technologies, the field of genomics has had an immense boost.

Application of these new technologies have rapidly become common practice in many research and diagnostic fields.

Sequencing whole genomes of known and new species are now routine experiments.

The GIGA genomics platform offers a wide range of services in bulk or single cell DNA/RNA analysis. Technologies ranges from Sanger sequencing to high throughput genotyping, high throughput sequencing and to long read sequencing.

Beside generating data, the platform also offers services in bioinformatics analysis.

[www.gigagenomics.uliege.be](http://www.gigagenomics.uliege.be)



## EXPERIENCED STAFF



We help you in setting up the experimental design



We produce sequencing libraries for many applications



We sequence on short or long read sequencing platforms



We generate QC reports on the results



We provide support/advice in the analysis of your results



We analyze your data in depth in close collaboration with you

## EQUIPMENT

**ABI 3700 Sanger sequencer**  
48 fragments sequenced up to 1800 bp

**2•Illumina MiSeq**  
25•10<sup>6</sup> fragments sequenced up to 600 bp

**2•Illumina NextSeq500**  
400•10<sup>6</sup> fragments sequenced up to 300 bp

**Illumina NovaSeq6000**  
20•10<sup>9</sup> fragments sequenced up to 300 bp

**Oxford Nanopore long read sequencer**  
10 to 20 Gbp of long reads >10kb

**2•Illumina iScan + autoloader**  
Cost efficient array genotyping, up to 2000 samples/week

**Chromium 10x Genomics**  
High throughput Single cell transcriptomics

**Computer cluster**  
552 cores and 4.8T ram

**Secured storage**  
1500T disk storage,  
1500T tape storage

## APPLICATIONS

### Data generation

De novo genome sequencing

Whole genome re-sequencing

Bulk transcriptome analysis

Single cell transcriptome analysis

Long read DNA/RNA sequencing

Cohort genotyping

Metagenomics

Amplicon sequencing

TCR repertoire sequencing

Ribosome profiling

### Data analysis

De novo assembly

Genome wide association analysis (GWAS)

Differential expression analysis

Single cell expression analysis

RNA velocity

Whole genome variant calling

## GIGA PLATFORMS



Genomics



Cell Imaging



Flow Cytometry



CRC in vivo Imaging



CRC Preclinical Imaging



Immunohistology



Proteomics



Viral Vectors



Mouse Facility



Zebrafish Facility

## CONTACTS

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+32 4 242 77 60

# SEQUENCERS



**MiSeq**  
540 Mb -15 Gb  
4 – 56 hours



**HiSeq**  
105 Gb - 1,5 Tb  
1 – 3,5 days



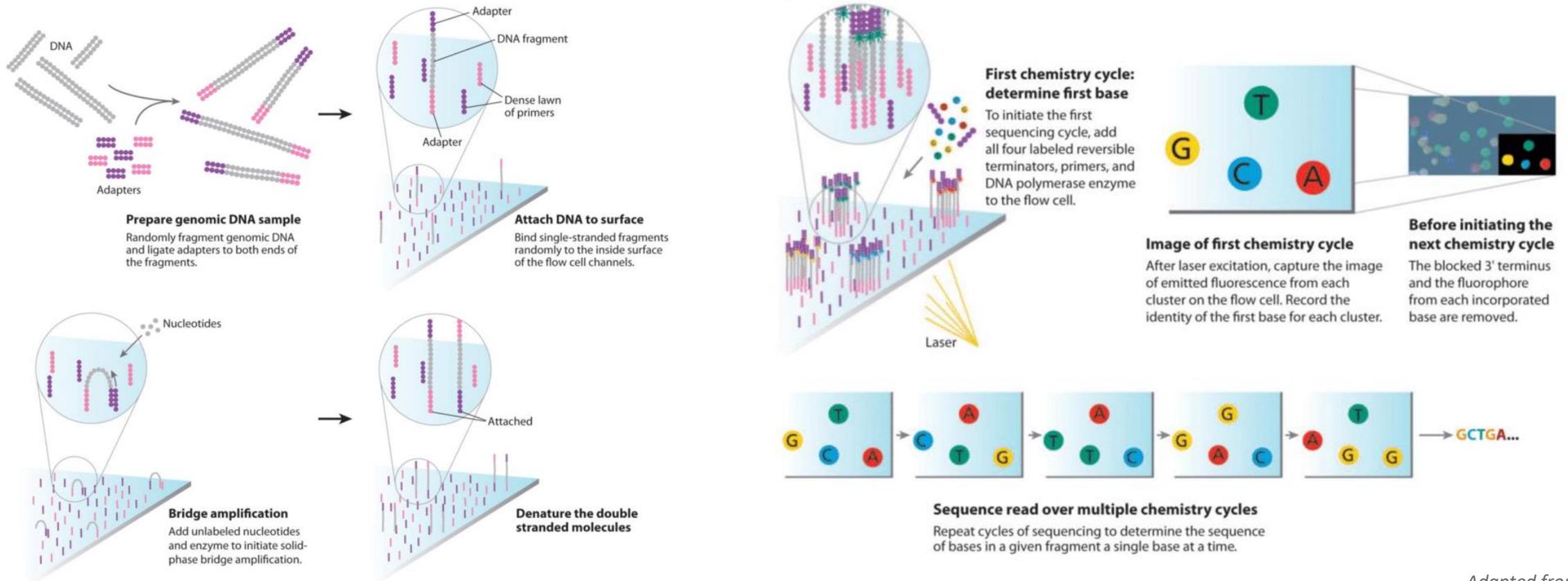
**NextSeq**  
16,25 Gb - 120 Gb  
11 – 29 hours



**NovaSeq**  
65 Gb – 3 Tb  
13 – 44 hours

*Adapted from Illumina*

# SEQUENCING (HTS/NGS)



*Adapted from Illumina*

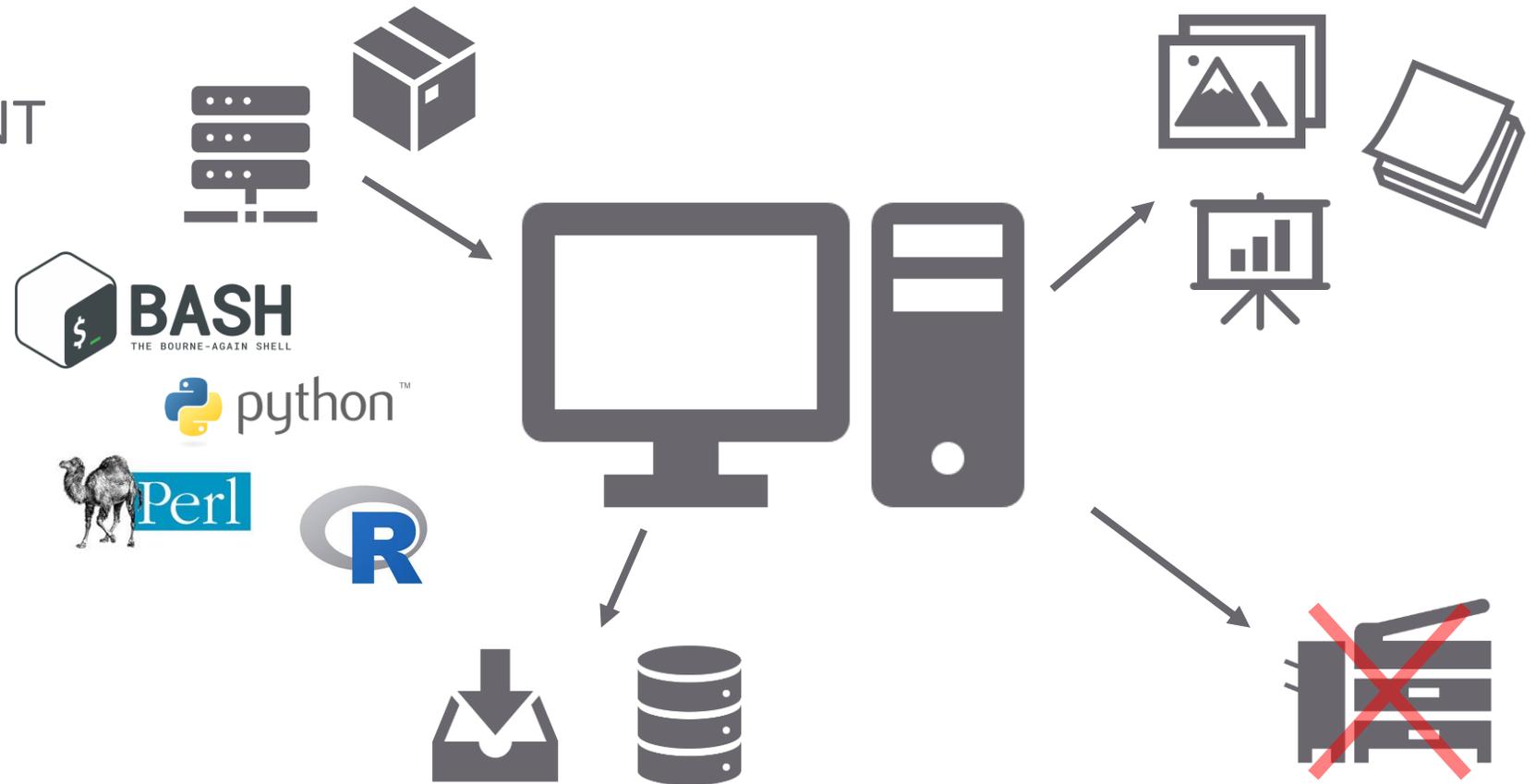
# COMPUTING

- DATA MANAGEMENT

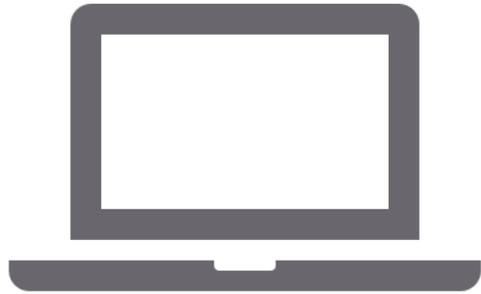
- DATA PROCESSING
- STORAGE
- RESULTS

- REPRODUCIBILITY

- PIPELINES
- CONTAINERS



# DATA MANAGEMENT



**LAPTOP**  
250Gb – 1Tb  
4-8 Go RAM  
4-8 CPUs



**LAB COMPUTER**  
1 - 5 Tb  
32 - 128 Go RAM  
8-16 CPUs



**CLUSTER/STORAGE**  
1.5 Pb  
256 Go RAM  
32 CPUs

# DATA MANAGEMENT

August 28, 2020



- 1.2 Tb
- 12.8 Billions of reads
- 612 samples
  - Multiple experiments
  - Unique combinations of indexes
- « Run »
- No storage on device



# DATA PROCESSING

- DEMULTIPLEXING

- Reads → Samples
- ~ 20M / sample
- « Fastq »
  - Identifier
  - Sequence
  - Separator
  - Quality score
    - Ascii +33

- + 1.2 Tb (2.4 Tb)



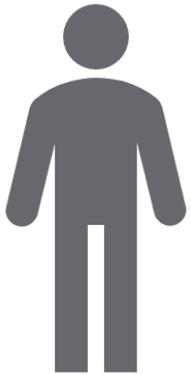
```
@A00801:49:H2VGKDSXY:2:1101:1624:1016 1:N:0:NGCTTAAG+TCGTGACC
CTTCTGGAGAGGAGTTCTCTGATATGAATTAAGGTTTTCCCTCTGTGCATGACCAGAAGAAGGTTTTATCTGTGCCACACTACTTTTCATTTCTGTTGCCAGTTGGTCCAATA
AATCAAAGATGNTTCAAACCTGGTCCAATAACAAGT
+
FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF
FFFFFFFFF
@A00801:49:H2VGKDSXY:2:1101:2022:1016 1:N:0:NGCTTAAG+TCGTGACC
TGACAAAAGATACCTCATTATGGGGAAATTGAGGAAGATACATATACAAGCACCCCAACCCATATTTAACATATTTGGCAATAACTCCCTCCATTCTCCCCCTCAATT
TCAAATAGTAGNTTTTAAAAAATTAAGACATGTC
+
FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF
:FF,FF
@A00801:49:H2VGKDSXY:2:1101:4137:1016 1:N:0:NGCTTAAG+TCGTGACC
TTTTTTGCCCTTTCAAGTGTTATTTTATACATTTTTTGTATTA AAAAGAAAAGCATAATTACCACAAATTACAAAGGACTAAAGCAGGACTAGAATAATGAATGAATCAC
TTCAGCTGGAANGCAGATACTCTCAATAATTAAT
+
FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF
FFFFF
```

Symbol	ASCII Code	Q-Score	Symbol	ASCII Code	Q-Score
!	33	0	6	54	21
"	34	1	7	55	22
#	35	2	8	56	23
\$	36	3	9	57	24
%	37	4	:	58	25
&	38	5	;	59	26
'	39	6	<	60	27
(	40	7	=	61	28
)	41	8	>	62	29
*	42	9	?	63	30
+	43	10	@	64	31
,	44	11	A	65	32
-	45	12	B	66	33
.	46	13	C	67	34
/	47	14	D	68	35
0	48	15	E	69	36
1	49	16	F	70	37
2	50	17	G	71	38
3	51	18	H	72	39
4	52	19	I	73	40
5	53	20			

Phred Quality Score	Probability of incorrect base call	Base call accuracy
10	1 in 10	90,0000%
20	1 in 100	99,0000%
30	1 in 1000	99,9000%
40	1 in 10,000	99,9900%
50	1 in 100,000	99,9990%
60	1 in 1,000,000	99,9999%

# RESEARCHERS





- 12 samples
- Fastq
- 52 Gb
- RNA-Seq

Transcriptomics  
« Gene Expression »

## WORKFLOW

- QC Sequencing
- Mapping
- Quantification
- QC Mapping/Quantification
  
- Downstream Analysis
  - Clustering
  - Differential Expression

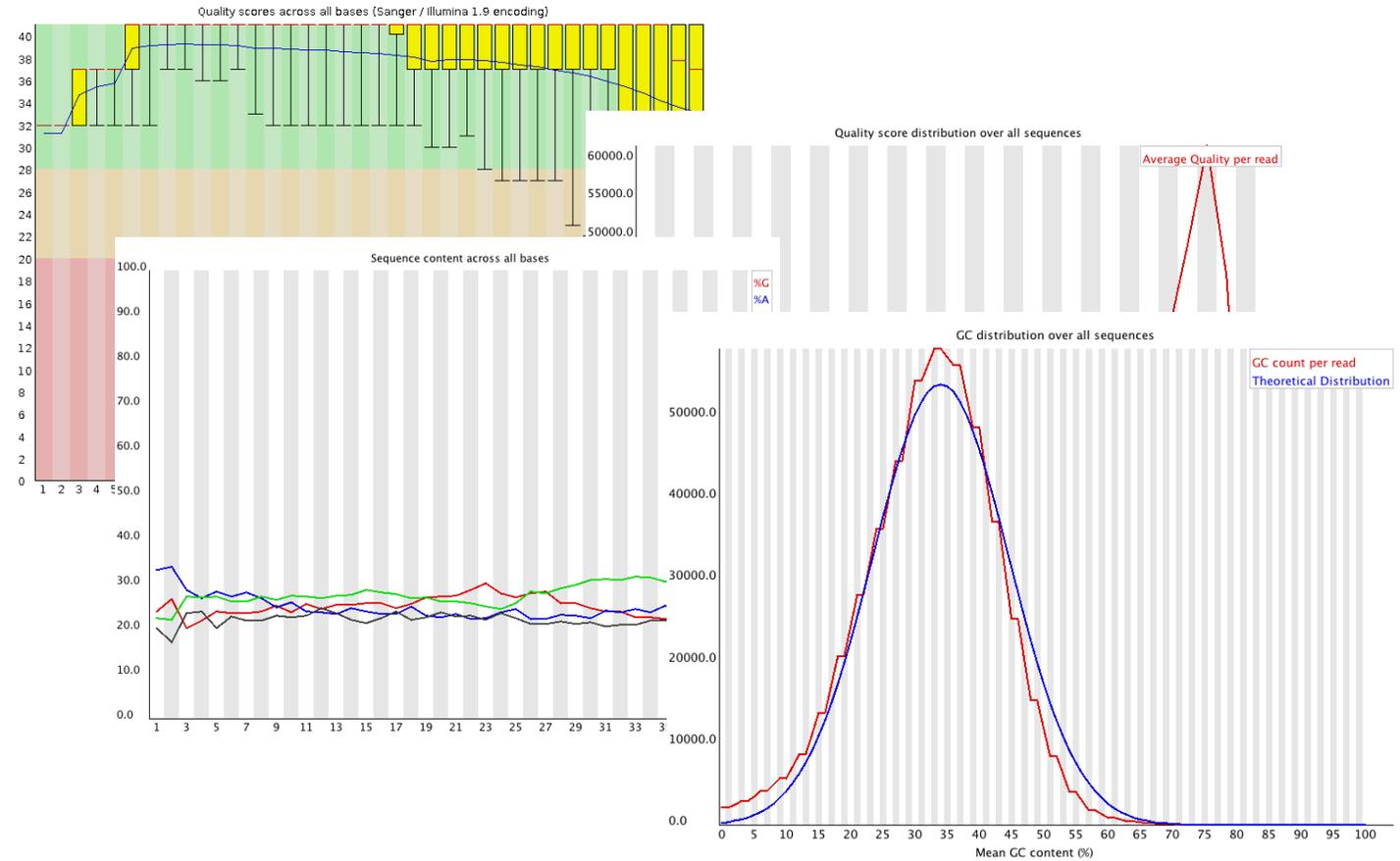
# QC SEQUENCING

Dr GIGA  
52 Gb



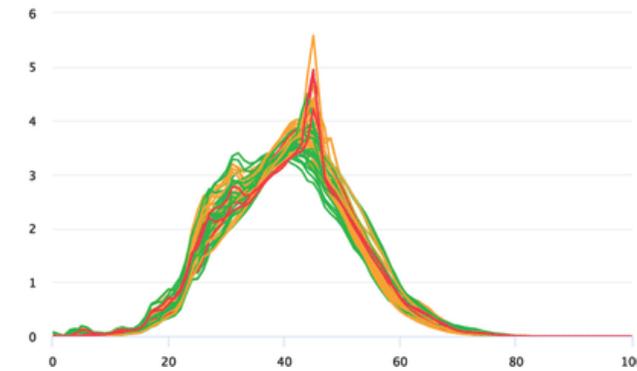
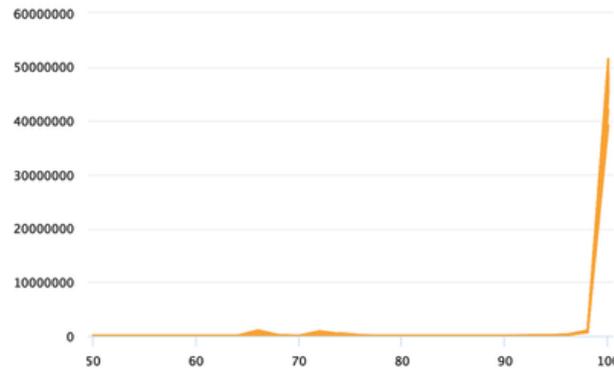
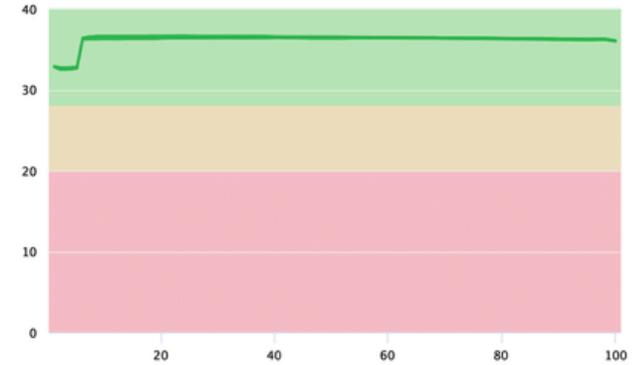
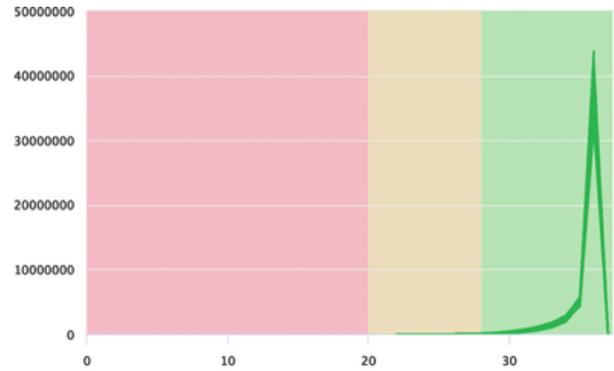
- Number of reads
- Base calling quality
- Sequence quality
- GC content
- Sequence length
- Duplication levels
- Adapter content
- Overrepresented sequences
- ...

< 1 Mb



# QC SEQUENCING

Dr GIGA  
52 Gb



# MAPPING

Dr GIGA  
52 Gb

- Alignment
- Origin of reads
- Reference
  - Genome sequence
  - Gene set
- Database
  - Ensembl, UCSC, ...

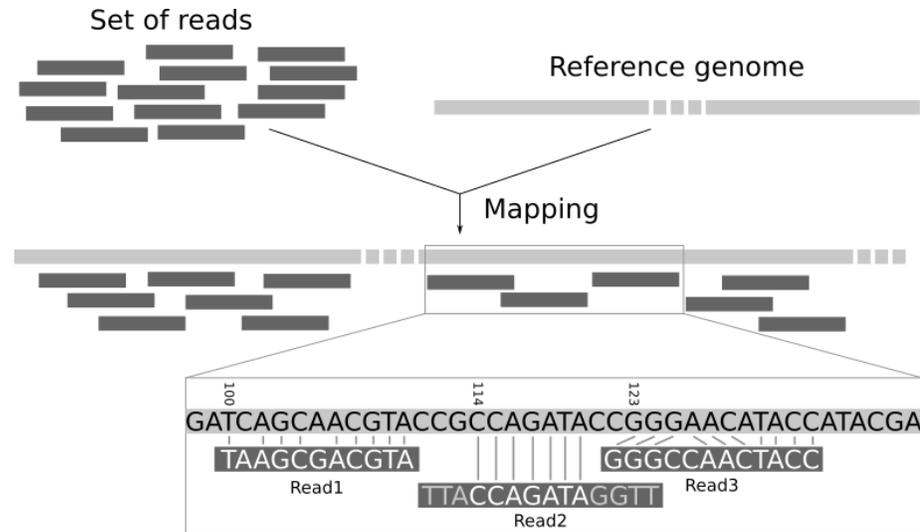




# MAPPING

Dr GIGA  
56 Gb

- Homo Sapiens
  - Genome (3.2 Gb)
  - Gene Set (1.2 Gb)
- Softwares
  - STAR
  - HISAT
  - ...
- High RAM/CPU's



# MAPPING

Dr GIGA  
86 Gb

- Genome Indexing
  - Quick queries
  - 20M reads



- High RAM/CPU



- H.Sapiens ~30 Gb



# MAPPING

Dr GIGA  
118 Gb

- SAM/BAM files (~ 32 Gb)
  - FLAG - Information
  - RNAME - Chromosome
  - POS – Location of 1st base
  - MAPQ – Quality score
  - CIGAR - Operations

Flag	Description
1	read is mapped
2	read is mapped as part of a pair
4	read is unmapped
8	mate is unmapped
16	read reverse strand
32	mate reverse strand
64	first in pair
128	second in pair
256	not primary alignment
512	read fails platform/vendor quality checks
1024	read is PCR or optical duplicate

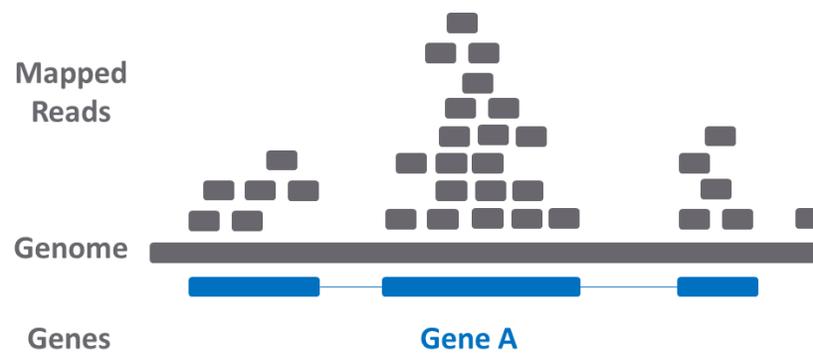
Paired-End

```

A00801:76:HGJCYDSXY:4:1544:20401:36699 99 1 3112677 255 150M = 3112770 244
CTAGGAGATAGTAGGGATTGGGAAGCAACTACTGAAAGGTCTGTGTCTTCTTTGTGGATGATAAAATATTCTGGAATTATATTGTATGCTAGGCGCACAACTTGTGACCATAGTACAGATATTCAACAGATAAATTTGTGTGCTATGA
F:FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF
NH:i:1 HI:i:1 AS:i:299 nM:i:0 RG:Z:SV2-CTRL2_NGS20-0393_AHGJCYDSXY_S241_L004_R1_001
    
```

# QUANTIFICATION (RNA)

- Gene Expression



	union	intersection_strict	intersection_nonempty
	gene_A	gene_A	gene_A
	gene_A	no_feature	gene_A
	gene_A	no_feature	gene_A
	gene_A	gene_A	gene_A
	gene_A	gene_A	gene_A
	ambiguous	gene_A	gene_A
	ambiguous	ambiguous	ambiguous

# QUANTIFICATION (RNA)

Dr GIGA  
118 Gb

- Gene Expression
- « Count matrix » (3.2 Mb)
- Major output



Each column is a sample

GENE ID	KD.2	KD.3	OE.1	OE.2	OE.3	IR.1	IR.2	IR.3
1/2-SBSRNA4	57	41	64	55	38	45	31	39
A1BG	71	40	100	81	41	77	58	40
A1BG-AS1	256	177	220	189	107	213	172	126
A1CF	0	1	1	0	0	0	0	0
A2LD1	146	81	138	125	52	91	80	50
A2M	10	9	2	5	2	9	8	4
A2ML1	3	2	6	5	2	2	1	0
A2MP1	0	0	2	1	3	0	2	1
A4GALT	56	37	107	118	65	49	52	37
A4GNT	0	0	0	0	1	0	0	0
AA06	0	0	0	0	0	0	0	0
AAA1	0	0	1	0	0	0	0	0
AAAS	2288	1363	1753	1727	835	1672	1389	1121
AACS	1586	923	951	967	484	938	771	635
AACSP1	1	1	3	0	1	1	1	3
AADAC	0	0	0	0	0	0	0	0
AADACL2	0	0	0	0	0	0	0	0
AADACL3	0	0	0	0	0	0	0	0
AADACL4	0	0	1	1	0	0	0	0
AADAT	856	539	593	576	359	567	521	416
AAGAB	4648	2550	2648	2356	1481	3265	2790	2118
AAK1	2310	1384	1869	1602	980	1675	1614	1108
AAMP	5198	3081	3179	3137	1721	4061	3304	2623
AANAT	7	7	12	12	4	6	2	7
AARS	5570	3323	4782	4580	2473	3953	3339	2666

Each row is a gene

# QC MAPPING/QUANTIFICATION

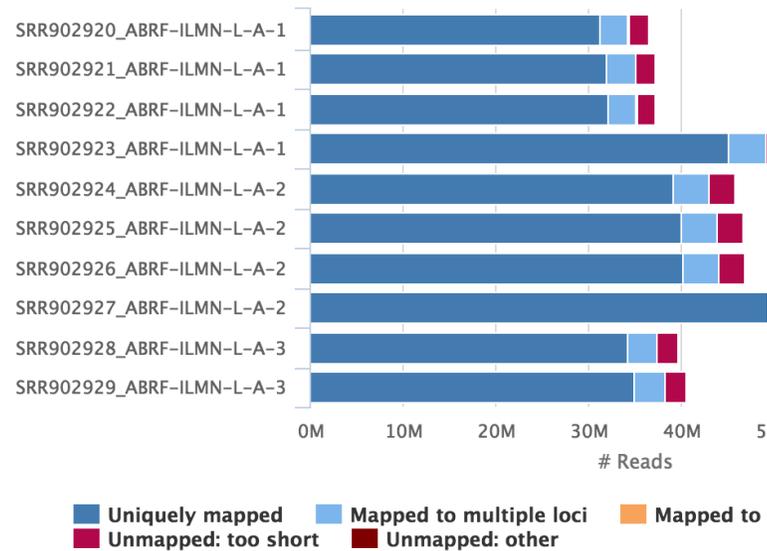


## General Statistics

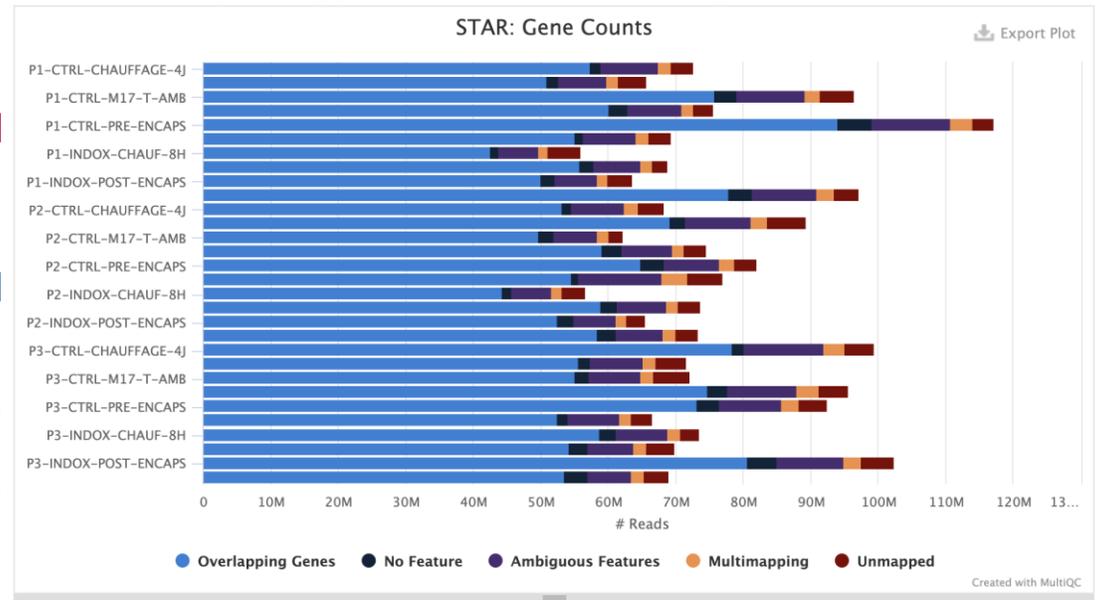
[Copy table](#)
[Configure Columns](#)
[Plot](#)
 Showing 8/8 rows and 8/10 columns.

Sample Name	% Assigned	M Assigned	% Aligned	M Aligned	% Trimmed	% Dups	% GC	M Seqs
SRR902920	97.8%	104.4	4.0%	97.8	4.0%	78.9%	51%	104.4
SRR902921	87.1%	92.0	3.5%	87.1	3.5%	77.2%	49%	92.0

### STAR Alignment Scores



### STAR: Gene Counts



# DOWNSTREAM ANALYSIS

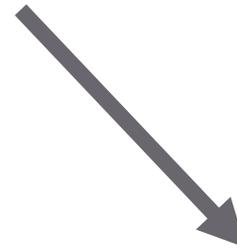
- Experimental Design

- R

- DESeq2
- EdgeR
- Voom
- ROTS
- Limma
- ...

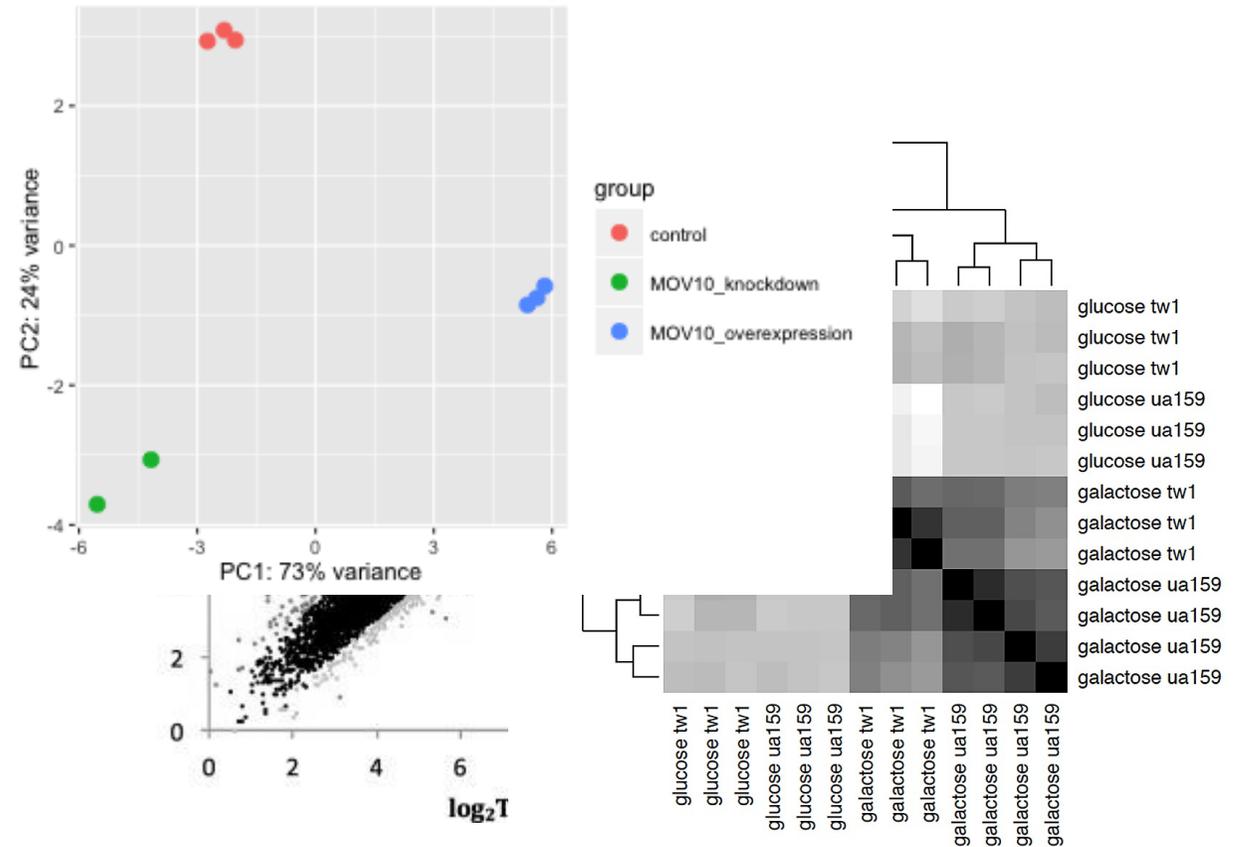


Counts



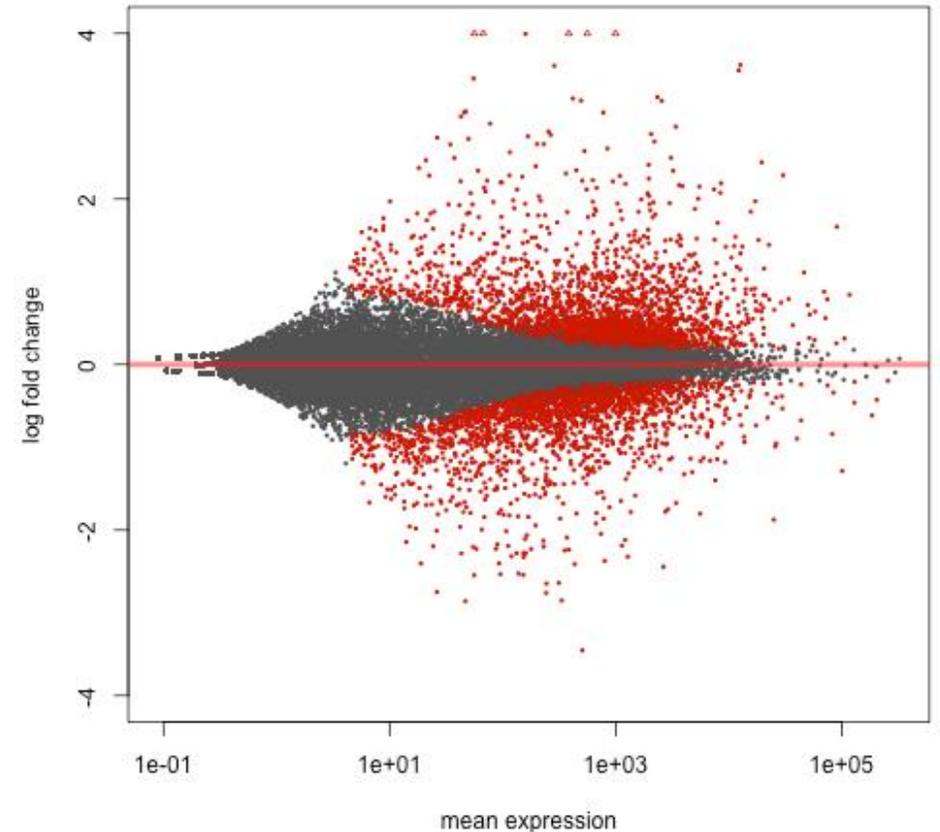
# DOWNSTREAM ANALYSIS

- Experimental Design
- Clustering
  - Sample correlation
  - Euclidian distance
  - Principal Component Analysis



# DOWNSTREAM ANALYSIS

- Experimental Design
- Clustering
  - Sample correlation
  - Euclidian distance
  - Principal Component Analysis
- Differential Expression Analysis
  - Pairwise comparisons
  - MA plot



# DOWNSTREAM ANALYSIS

- Experimental Design
- Clustering
  - Sample correlation
  - Euclidian distance
  - Principal Component Analysis
- Differential Expression Analysis
  - Pairwise comparisons
  - MA plot
  - DE genes

	baseMean	log2FoldChange	lfcSE	stat	pvalue	padj
PAX5	1531,91362	6,1280938	0.14505696	28,091789	1,23E-173	1,59E-169
SOX9	348,04912	3,5537120	0.15748166	20,475861	3,53E-93	2,27E-89
PDX1	830,75570	-1,8973788	0.12438094	-15,203018	3,38E-52	1,45E-48
ISL1	655,25202	-1,9729198	0.13344796	-14,715372	5,14E-49	1,65E-45
ARX	526,74210	2,2554297	0.15754888	14,227414	6,19E-46	1,59E-42

## Summary

out of 21769 with nonzero total read count  
adjusted p-value < 0.1

LFC > 0 (up) : 985, 4.5%

LFC < 0 (down) : 929, 4.3%

outliers [1] : 0, 0%

low counts [2] : 8914, 41%

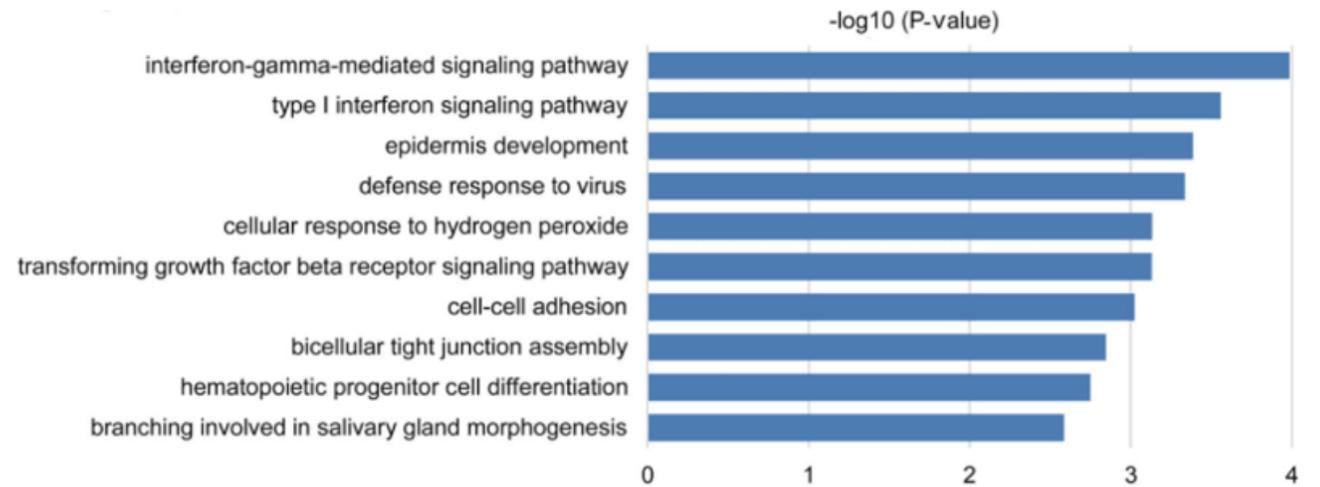
(mean count < 8)

[1] see 'cooksCutoff' argument of ?results

[2] see 'independentFiltering' argument of ?results

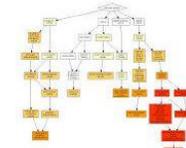
# DOWNSTREAM ANALYSIS

- Biological meaning
- Gene ontology / Gene Set Enrichment Analysis
  - GSEA
  - Enrichr
  - GOrilla
  - PANTHER
  - ...

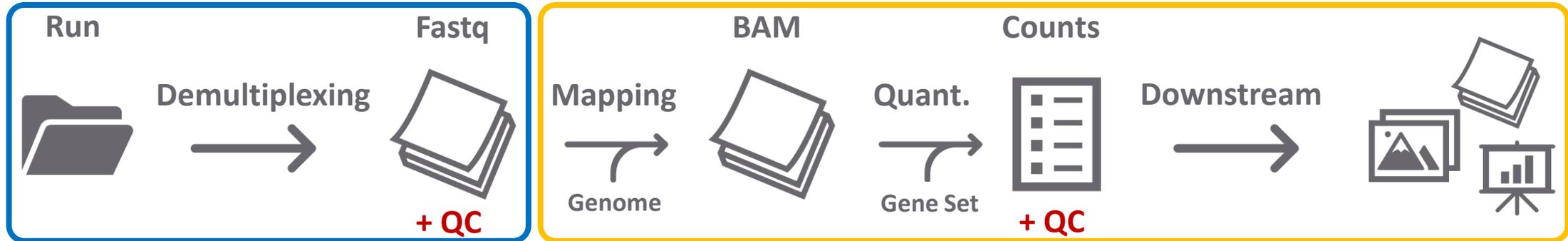


**GORILLA**

*Gene Ontology enRIchment anaLysis and visuaLiZAtion tool*



# SUMMARY



# REPRODUCIBILITY

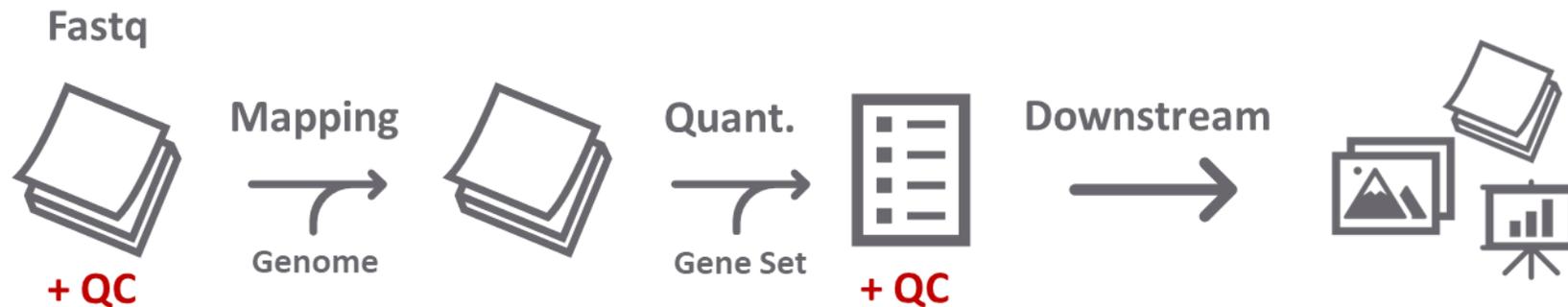
PIPELINES & CONTAINERS

# REPRODUCIBILITY

- Pipelines
  - Set of successive actions
    - Softwares
    - Parameters
    - References

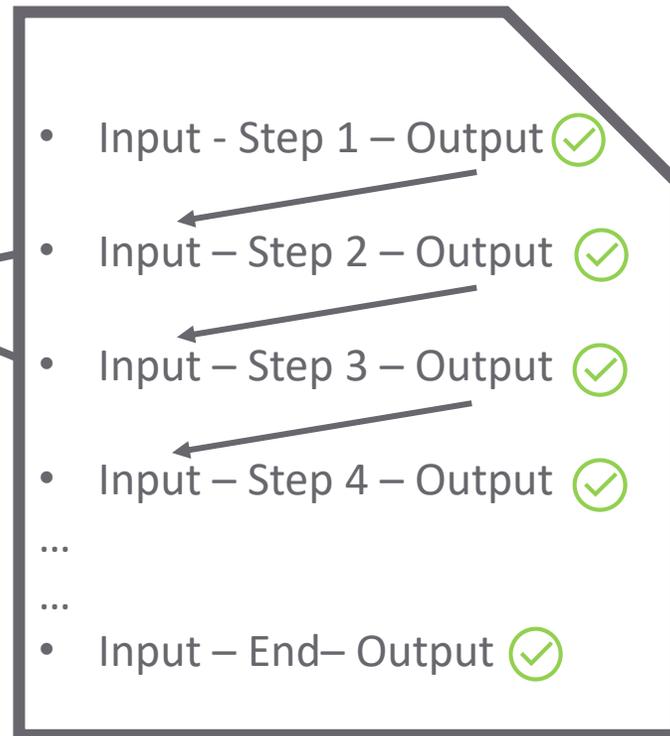
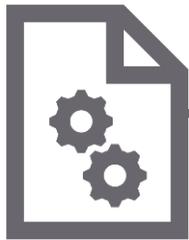
nextflow

Snakemake



# REPRODUCIBILITY

- Pipelines
  - Scripts

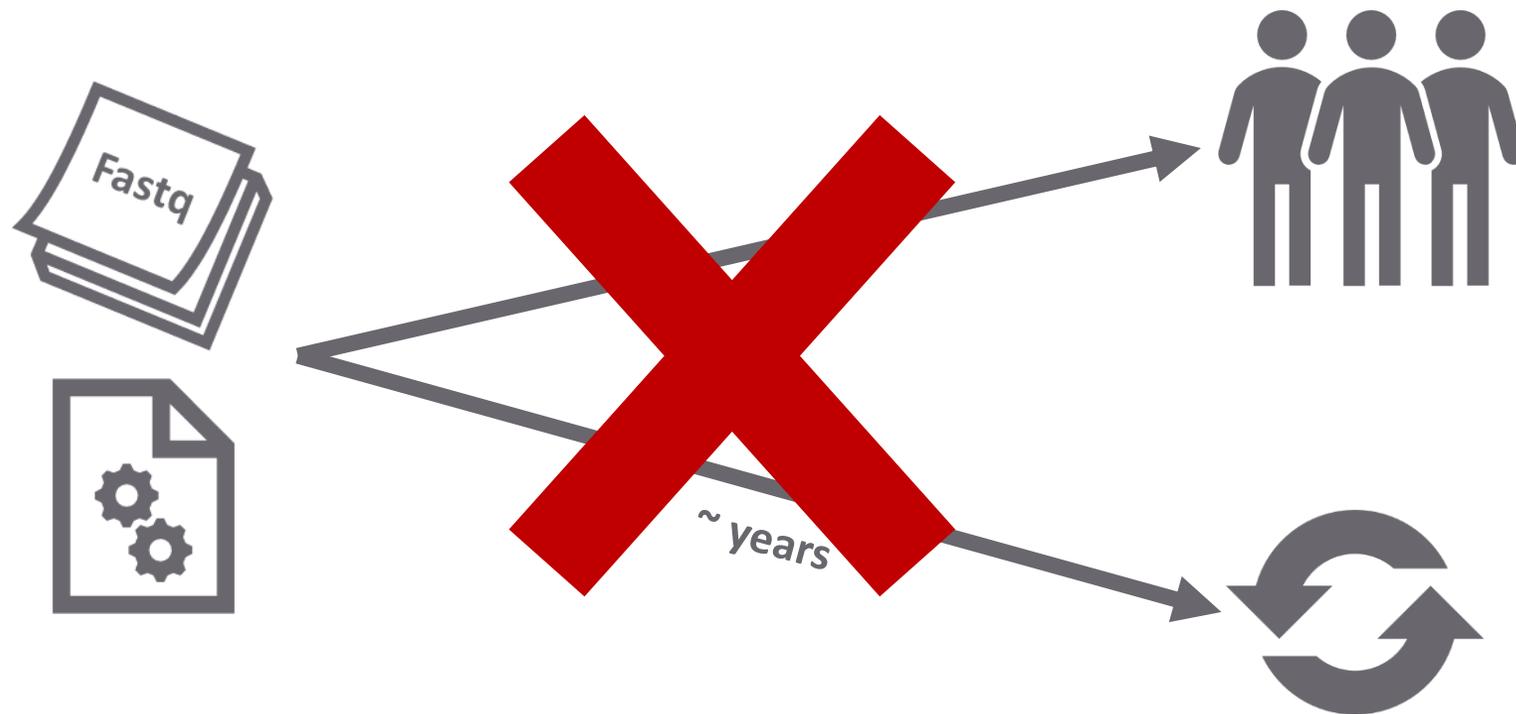


**nextflow**

Snakemake

**nf-core** 

# REPRODUCIBILITY



# REPRODUCIBILITY

- Variability

- Updates / Versioning

- Softwares

- References

- Compatibility

- Format

- Knowledge

STAR 2.7.5b - 2020/08/01  
STAR 2.7.5c - 2020/08/16  
STAR 2.7.6a - 2020/09/19

**List of currently available archives**

- [Ensembl GRCh37](#): Full Feb 2014 archive with BLAST, VEP and BioMart
- [Ensembl 101: Aug 2020](#) - this site
- [Ensembl 100: Apr 2020](#)
- [Ensembl 99: Jan 2020](#)
- [Ensembl 98: Sep 2019](#)
- [Ensembl 97: Jul 2019](#)
- [Ensembl 96: Apr 2019](#)
- [Ensembl 95: Jan 2019](#)
- [Ensembl 94: Oct 2018](#)
- [Ensembl 93: Jul 2018](#)
- [Ensembl 92: Apr 2018](#)
- [Ensembl 91: Dec 2017](#)

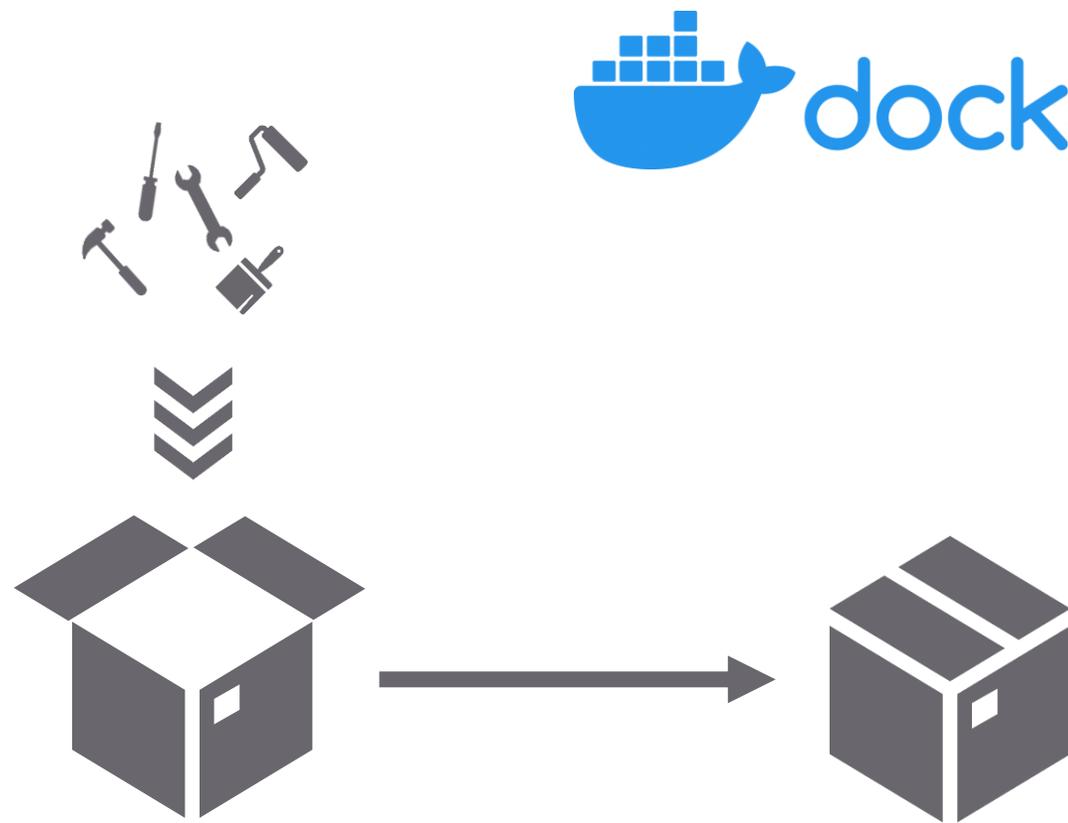
# REPRODUCIBILITY

- CONTAINERS

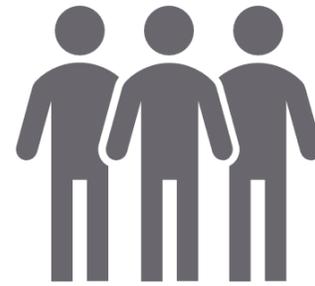
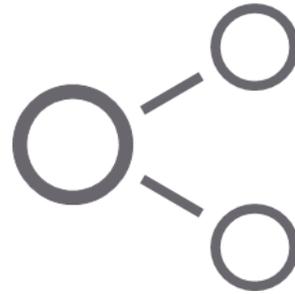
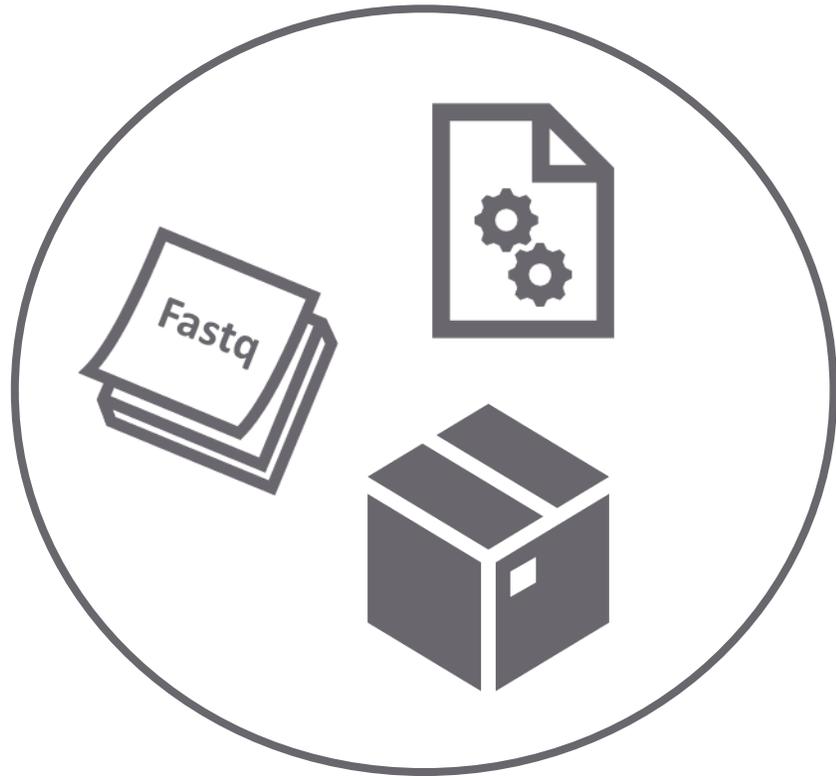
- Docker
- Singularity

- Softwares

- Versions



# REPRODUCIBILITY



# DATA DEPOSITORY

- Gene Expression Omnibus (NCBI)
- ArrayExpress (EMBL-EBI)



THANK YOU FOR YOUR ATTENTION